



Contents lists available at ScienceDirect

Life Sciences

journal homepage: www.elsevier.com/locate/lifescie



Current understanding of structural and molecular changes in diabetic cardiomyopathy

Md Sayeed Akhtar^{a,*}, Sirajudeen S. Alavudeen^a, Asif Raza^b, Mohammad Tarique Imam^c, Ziad Saeed Almalki^c, Fauzia Tabassum^{d,e}, Mir Javid Iqbal^f

^a Department of Clinical Pharmacy, College of Pharmacy, King Khalid University, Al-Fara, Abha 62223, Saudi Arabia

^b Department of Pharmacology, Penn State Cancer Institute, CH72, Penn State College of Medicine, Penn State Milton S. Hershey Medical Center, 500 University Drive, Hershey, PA 17033, USA

^c Department of Clinical Pharmacy, College of Pharmacy, Prince Sattam Bin Abdulaziz University, Al-Kharj 16273, Saudi Arabia

^d Department of Pharmacology, College of Dentistry and Pharmacy, Buraydah Private College, Al Qassim 51418, Saudi Arabia

^e Department of Pharmacology, Vision College, Ishbilia, Riyadh 13226-3830, Saudi Arabia

^f Department of Pharmaceutical Sciences, Northeastern University, Boston, MA 02115, USA

ARTICLE INFO

Keywords:

Understanding
Diabetes mellitus
Cardiomyopathy
Pathophysiology
Therapy

ABSTRACT

Diabetic Mellitus has been characterized as the most prevalent disease throughout the globe associated with the serious morbidity and mortality of vital organs. Cardiomyopathy is the major leading complication of diabetes and within this, myocardial dysfunction or failure is the leading cause of the emergency hospital admission. The review is aimed to comprehend the perspectives associated with diabetes-induced cardiovascular complications. The data was collected from several electronic databases such as Google Scholar, Science Direct, ACS publication, PubMed, Springer, etc. using the keywords such as diabetes and its associated complication, the prevalence of diabetes, the anatomical and physiological mechanism of diabetes-induced cardiomyopathy, the molecular mechanism of diabetes-induced cardiomyopathy, oxidative stress, and inflammatory stress, etc. The collected scientific data was screened by different experts based on the inclusion and exclusion criteria of the study. This review findings revealed that diabetes is associated with inefficient substrate utilization, inability to increase glucose metabolism and advanced glycation end products within the diabetic heart resulting in mitochondrial uncoupling, glucotoxicity, lipotoxicity, and initially subclinical cardiac dysfunction and finally in overt heart failure. Furthermore, several factors such as hypertension, overexpression of renin angiotensin system, hypertrophic obesity, etc. have been seen as majorly associated with cardiomyopathy. The molecular examination showed biochemical disability and generation of the varieties of free radicals and inflammatory cytokines and becomes are the substantial causes of cardiomyopathy. This review provides a better understanding of the involved pathophysiology and offers an open platform for discussing and targeting therapy in alleviating diabetes-induced early heart failure or cardiomyopathy.

1. Introduction

A significant public health issue that affects the entire world's population is diabetes mellitus. The steady and silent impact of diabetes mellitus (DM) in the development of functional cardiac disorder is well evident. Hyperglycemia and insulin resistance (IR) are two features of type 2 diabetes (T2DM), a metabolic condition that is chronic and increases the risk of heart failure (HF) [1]. Even in the absence of myocardial ischemic or microvascular atherosclerotic disease, long-term diabetes causes structural and functional changes in the myocardium

that cause HF to develop and progress. These changes are a direct result of abnormal myocardial metabolism and IR [2]. In addition to increasing the risk of HF hospitalization, all-cause mortality, and CV mortality, the presence of diabetes with HF is a poor prognostic indicator. For instance, epidemiologic studies showed that individuals with HF and diabetes had a 50–90 % increased risk of CV mortality, regardless of the HF phenotype [3]. Many studies reported the risk of HF due to DM independent of hypertension (HTN), coronary heart disease (CHD), and valvular heart disease (VHD) [4,5]. The idea of diabetic cardiomyopathy (DCM) was first proposed by Rubler et al after post-mortem studies in diabetic

* Corresponding author at: Department of Clinical Pharmacy, College of Pharmacy, King Khalid University, Al-Fara, Abha 62223, Saudi Arabia.
E-mail address: mdhsuain@kku.edu.sa (M.S. Akhtar).

patients with cardiac failure. They excluded alcohol, hypertension, coronary and structural heart disease as possible etiologies and considered only DM as the etiologic factor [2]. Moreover, a relative increase in left ventricular (LV) thickness, LV mass, and other related structural abnormalities has been also observed in the diabetic heart that develops LV dysfunction [6]. These changes are also accompanied due to microvascular modifications followed by myocardial fibrosis and remodeling in the extracellular matrix [7]. Thus, in-depth understanding of molecular changes involved in pathophysiological alterations must be looked at for managing the potential loss of left ventricular dysfunction in a patient with DM.

2. Global burden

The Framingham Heart Study reported an increased incidence of HF in diabetic patients in comparison to the age-matched population independent of other concomitant diseases like obesity, hypertension, dyslipidemia, and coronary heart disease [8]. International diabetic federation reported that 463 million people are currently suffering from DM and by 2045, the number of diabetic cases will rise to more than 700 million. However, almost 232 million cases of DM are currently undiagnosed. Moreover, 79 % of the total diabetic population is living in low- and middle-income countries, where the prevalence is alarming because of low socioeconomic status [9]. According to a recent temporal analysis of a sizable UK cohort, the prevalence of DM in HF has significantly increased (18 % in the early 2000s against 26 % in recent years) [10].

3. Diabetic cardiomyopathy

Among cardiovascular diseases, DCM is the utmost cause of mortality and debility among the diabetic population other than cardiovascular diseases (CVDs) such as myocardial angina, myocardial infarction (MI), stroke, arterial disorder, and congestive HF [11]. Hypertension, hypercholesterolemia, persistent hyperglycemia, and other risk factors like a sedentary lifestyle have been observed to be significantly involved in raising the risk of cardiovascular complications in diabetics [12]. Data from both clinical and experimental studies represents that DCM is a major complication that contributes to HF other than atherosclerosis, high blood pressure, and other related complications [13]. DCM consists of earlier short-term, physiological adaptation to metabolic changes, and thereafter degenerative changes for which the myocardium has only limited capacity to heal [12,14]. There are many factors (Fig. 1) such as prescription drugs, metabolic characteristics, and lipid status that may also influence the progression of DCM [15]. Therefore, the treatment during the early stages of DM can potentially delay or obstruct the progression of other consequences [16]. The different stages of DCM have been mentioned in Table 1.

4. Structural changes in diabetic cardiomyopathy

4.1. Microvascular dysfunction

Coronary microcirculation regulates the coronary blood flow and maintains the oxygen demand in cardiac tissue. Microvascular dysfunction especially in the coronary artery causes impaired

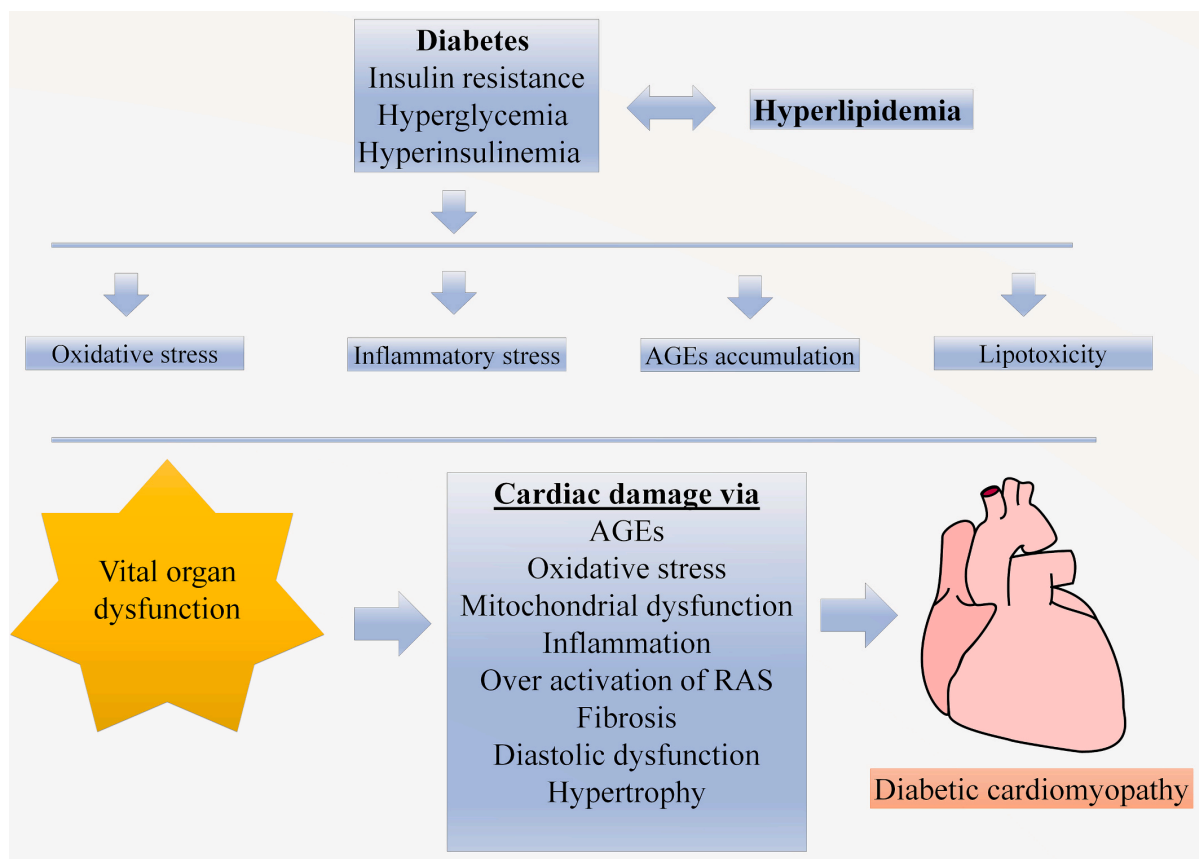


Fig. 1. Physiological and biochemical changes in diabetic cardiomyopathy.

Table 1
Stages of diabetic cardiomyopathy.

Stages	Molecular changes	Functional changes	Structural changes	Reference
Early stage	<ul style="list-style-type: none"> • Depletion of GLUT4 • Increased FFA • L-carnitine deficiency • Imbalance in Ca^{2+} homeostasis • Insulin resistance 	No overt functional abnormalities or possible overt diastolic dysfunction or changes in ejection fraction	Normal LV size with no sign of hypertrophy	[17–19]
Middle stage	<ul style="list-style-type: none"> • Increased AT II • Reduced IGF-I • Increased TGFβ1 • Mild CAN 	Abnormal diastolic dysfunction and normal or slightly decreased ejection fraction	Slightly increased LV mass, wall thickness, and evidence of hypertrophy	[20–22]
Last stage	<ul style="list-style-type: none"> • Microvascular changes • Hypertension • CAD • Severe CAN 	Abnormal diastolic dysfunction and ejection fraction	Significantly increased LV size, wall thickness, and well evidence of hypertrophy	[21–23]

GLUT-4: Glucose transporter-4, FFA: Free Fatty Acid, ATII: Angiotensin-II, IGF-I: Insulin-Like Growth Factor-1, TGF-β: Tissue Growth Factor, CAN: Cardiovascular Autonomic Neuropathy, CAD-Coronary Artery Disease, LV-Left Ventricle.

myocardial reserve volume and involves functional alteration of the myocardium [24]. Persistent hyperglycemia causes increased advanced glycation end products (AGEs) production concurrent to decreased production of NO in endothelial cells of coronary artery and this contributes to the development of DCM and subsequent CVD-related untoward events [25].

4.2. Interstitial fibrosis

DCM is characterized by interstitial and perivascular fibrosis. Cardiac biopsy showed that significant increase in collagen deposition in between intramural vessels and in an around myofibers in heart of diabetic patients [26]. A major elevation in collagen type III was reported after endomyocardial biopsies of the T2DM cases without any major CAD and hypertension [27]. This leads to myocardial fibrosis and cardio dynamic changes in the diabetic heart. The pathophysiology of the myocardial fibrosis of a diabetic heart includes a raised TGFβ1 receptor II, plasma MMP1, TIMP metalloproteinase inhibitor 1 (TIMP1), amino-terminal pro-peptide of type I and type III procollagen [28].

4.3. Cell death

Necrosis and apoptotic cell death are observed in biopsies of diabetic myocardium having concurrent HF [29]. Activated myocardial apoptosis due to leptin deficiency plays a major role in causing cell death through Rac1 mediated raised Reduced nicotinamide adenine dinucleotide phosphate (NADPH) and mitochondrial-derived ROS in diabetic myocardium [30]. Activation of the RAAS is linked with elevated oxidative stress, programmed cell death, and necrosis in myocardial and endothelial cells of cardiomyocytes in diabetic patients and also in end-stage HF. This represents another potential mechanism for cell death [31].

5. Physiological alterations in diabetic cardiomyopathy

5.1. Autonomic neuropathy

Chronic DM causes CAN that ultimately affects heart rate control, vascular hemodynamics, and cardiac morphology as well. The overstimulation of the sympathetic nervous system and suppression of parasympathetic nervous system activity is also well evident in DCM and other cardiac disorders [32].

5.2. Left ventricular hypertrophy

LV hypertrophy is characterized by an increased myocardial mass of the left ventricle, a major risk factor associated with HF that downs myocardial performance [33]. In the Framingham study, a significant increase in LV wall thickness was observed only in women with DM [34]. In a multiethnic subject with T2DM, the chances of increased LV mass are much greater after adjusting different covariates even hypertension [35]. In fact, in this same population, increased LV mass was observed only in diabetic cases, but not in cases with altered fasting blood glucose (BG) or impaired tolerance to glucose [36]. This indicates that the changes in cardiac geometry develop only after long-term steady-state hyperglycemia with or without obesity.

5.3. Systolic dysfunction

Systolic dysfunction develops in a later stage and usually starts after diastolic dysfunction. Compromised systolic dysfunction is often not detected using standard 2-dimensional echocardiography techniques. But, tissue Doppler strain analysis and peak systolic velocity calculation, and subtle systolic function have been described in around 24 % of randomly selected cases having DM, excluding subjects' cases having CAD or LVH [37].

5.4. Diastolic dysfunction

DCM in humans is identified by diastolic dysfunction. The tissue Doppler analysis reveals that 40–75 % of cases have diastolic dysfunction in both types of diabetic cases, independent of coronary artery disease. Cardiographic markers for diastolic dysfunction like E/E' and E/A ratios were abnormal in T2DM cases [38]. Moreover, diastolic dysfunctions have an interrelation with raised cardiac triglyceride levels concurrent with impaired Ca^{2+} ion reuptake [39].

5.5. Impaired contractile reserve

The contractile reserve is the most independent index and is calculated as the variation between myocardial contractility at rest and at stress either by exercise or by inotropic agents such as dobutamine [40]. Recent studies about DCM show multiple mechanisms that cause diastolic and systolic impairment with concurrent reduced contractile reserve. These mechanisms include accumulation of AGEs [41], adipokines [42], impairment of myocardial insulin signaling [43], altered calcium homeostasis [44], and lipotoxicity [45] that play a major role in developing reduced contractile reserve in DCM. Fig. 1 depicted the biochemical and structural changes in diabetic cardiomyopathy.

6. Molecular alterations underlying diabetic cardiomyopathy

Fig. 2 represents potential molecular abnormalities in the development of diabetic cardiomyopathy. We will present summaries of the molecular pathways that have been postulated to contribute to the development of diabetic cardiomyopathy in the following subsections, with an emphasis on both established and novel or developing mechanisms (summarised in Table 2). Many of these topics demand a thorough, impartial assessment. Due to space restrictions, we want to give

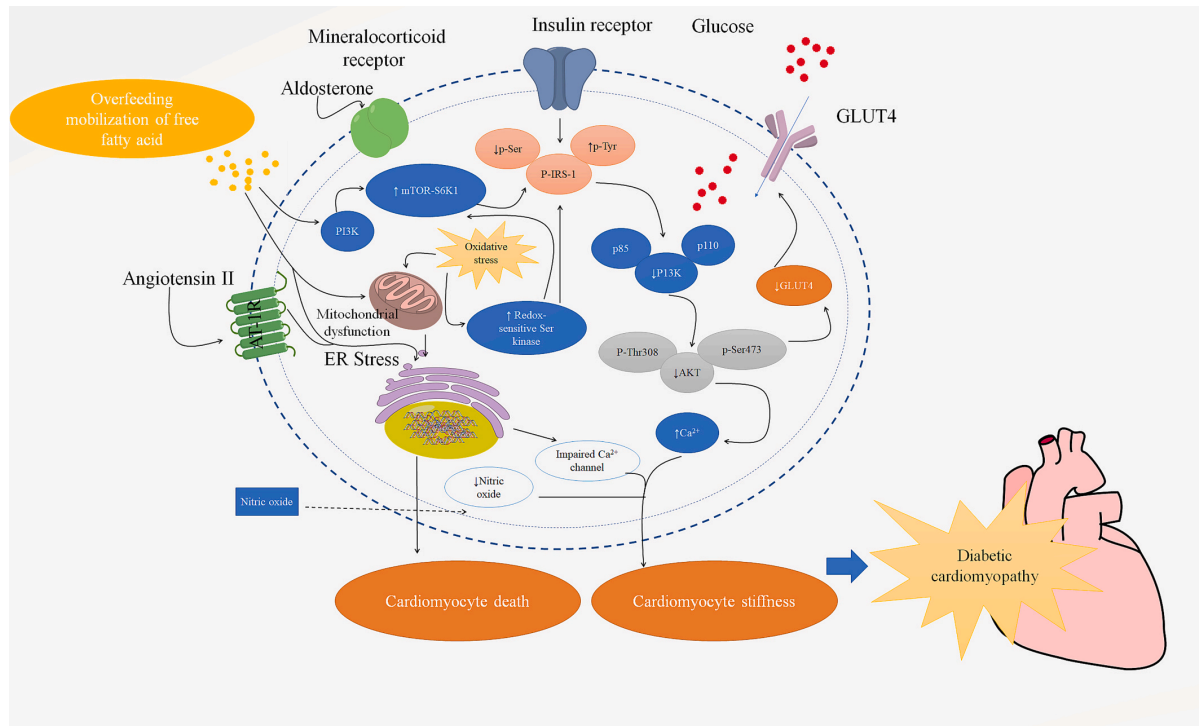


Fig. 2. Potential molecular abnormalities in the development of diabetic cardiomyopathy.

the reader an overview of the intricate pathophysiology of diabetic cardiomyopathy as a starting point for future in-depth investigation. In addition, though they are discussed separately, it is significant to realize that many of these mechanisms are closely related. The legend accompanying Fig. 2 includes examples of these interactions.

6.1. Myocardial lipotoxicity

T2DM is often associated with obesity which leads to myocardial lipotoxicity followed by cell death [46]. Regan and coworkers identified lipofuscin deposits, a brown lipid containing pigment granules in transmural LV biopsies obtained from patients with T2DM concurrent to raised myocardial triglycerides (TGs) and cholesterol [47]. Similarly, the staining of cardiac sections of a failing heart independent of ischemia with oil red-o staining showed a raised lipid deposition provoked by persistent hyperglycemia [45]. Hyperglycemia, obesity, insulin resistance, and impaired glucose tolerance (IGT) are related to raising intramyocardial lipids, independent of circulating triglycerides level [48]. However, raised myocardial triglycerides and their deposition is linked to diastolic but not systolic dysfunction [39]. Lipid-induced cell death might be an important contributor to developing DCM via activating ceramide biosynthesis [49]. In addition to this, alteration in the phospholipid makeup of the endoplasmic reticulum (ER) membrane has also been reported to cause swelling and trauma to ER [50,51].

6.2. Oxidative and nitrosative stress

Oxidative stress acts as a major contributor to the progression of DCM [52]. The mechanism behind the generation of ROS in the myocardium is still controversial probably due to lipid overload and vice versa.

Cellular sources of ROS and/or RNS generation within the heart include cardiac myocytes, endothelial cells, and neutrophils. A major

part of cellular ROS is produced in mitochondria and enzymatic systems that further generate the ROS in the cytosol mediated by NADPH oxidase [53]. These ROS and/or RNS initiate cellular damage through several mechanisms like oxidation, interference with NO, and modulation of detrimental intracellular signaling pathways such as increased expression of myocardial myosin heavy chain gene expression. This leads to cardiac dysfunction by direct damage to proteins and deoxyribonucleic acid (DNA), as well as by inducing apoptosis that led to cellular death [54]. Moreover, loss of the antioxidant defense system could also stimulate several other responses including activation of matrix metalloproteinase (MMP) to alter the architecture of the extracellular matrix and initiate cardiomyocyte hypertrophy that triggers ventricular remodeling [55]. Furthermore, Nrf2 (leucine zipper protein) activates the expression of hemoxygenase, an antioxidant protein in response to oxidative stress. However, persistent hyperglycemia and insulin resistance suppress Nrf2 expression and reduce their activity via Erk 1/2-mediated pathway. This further promotes oxidative stress and indirectly develops lipid accumulation, inflammation, fibrosis, and associated cardiac dysfunction in DCM [56].

6.3. Calcium signaling and ryanodine receptor

The calcium signaling system is the major pathway involved in activating the contractile system of cardiac myocytes through the ryanodine receptor (RyR), sarcoplasmic reticulum, and eNOS [57]. Moreover, RyRs play a major role in the cyclic rise and fall in intracellular Ca^{2+} ion of Ca^{2+} ions from the sarcoplasmic reticulum (SR) required for normal cardiac functions. DCM directly affects the ryanodine receptor complex and causes both reduction in ryanodine sensitivity to Ca^{2+} as well as a reduction in receptor volume that causes impaired myocardial contractility in DCM [33,58]. Apart from this, extremely important myocyte-specific enhancer factor 2C (MEF2C) downregulation is associated with the compromised activity of both sarcoplasmic endoplasmic

Table 2
Potential molecular abnormalities in the development of diabetic cardiomyopathy.

Molecular abnormalities	Consequence in the pathogenesis of diabetic Cardiomyopathy	References
Hyperglycaemia	Increased AGEs and ROS synthesis, Deactivation of NOS, myocardial collagen accumulation with fibrosis.	[13,22,121]
Myocardial lipotoxicity	Altered glycolysis, pyruvate oxidation, lactate causes apoptosis, perturbation of cardiac tissue bioenergetics with contraction/relaxation coupling. Stimulation of DAG/PKC signal transduction pathway causes decreased tissue blood flow, raised vascular permeability, changes in neovascularization and increased extracellular matrix deposition.	[46,122]
PKC	Cardiomyocyte hypertrophy and apoptosis	[123,124]
RAAS	Myofibroblast growth with interstitial and focal perivascular accumulation of collagen.	[63,125]
Aldosterone activation	Up-regulation of HIF-1 α through hypoxia/ROS stimulates angiopoietin, PGF, PDGF- β and VEGF. In diabetes, VEGF and its receptors, VEGF-R1 and VEGF-R2, reduced significantly, leads impairment of angiogenesis.	[64,126,127]
HIF-1/VEGF	Altered endothelial NO synthesis with raised vasoconstrictor PGs, glycated proteins, endothelium adhesion molecules and platelet and vascular growth factors enhance vasomotor tone and vascular permeability and limit growth and remodeling.	[128,129]
Endothelial dysfunction	Reduction in ryanodine receptor and its sensitivity to Ca ²⁺ ion. Reduced MEF2C activity followed by SECRC2a and NCX-1 downregulation.	[130–133]
Ca ²⁺ ion signaling and Raynodine receptor	Activation neutrophils, mast cells, dendritic cells, macrophages, eosinophils, nuclear transcription factor, NF κ B regulated proinflammatory cytokines like TNF α , interleukins MCP1, ICAM1, and VCAM1. NLRP-3 regulated inflammation and cellular death via NF κ B and TXNIP	[57,59]
Inflammatory cytokines	Reduced PI3K/Akt signaling initiates myocardial insulin resistance and autophagy, AMPK activation inhibits MTOR and activates autophagy, SIRT1/3 downregulation stresses the tissues, and FOXOs activation directly activates the autophagy protein.	[61,62,134]
Autophagy	transcriptional repression of GLUT4 by down regulation MEF2C. Activation of transcriptional enzymes via PPAR α involved in β oxidation. PDH overexpression suppresses the glucose oxidation and PPAR- α target gene mCPT1 activated lipotoxicity.	[77,135,136]
Altered substrate utilization and mitochondrial bioenergetics	miRNA-143, miRNA-181, miRNA-103, miRNA-107 and miRNA-802 regulates systemic glucose metabolism and insulin sensitivity. Dysregulation of the	[68]
miRNA		[80,86,137–139]

Table 2 (continued)

Molecular abnormalities	Consequence in the pathogenesis of diabetic Cardiomyopathy	References
	miRNA biogenesis via cardiomyocyte-specific deletion of dicer progresses to cardiomyopathy and HF. miRNA-133 induces fibrosis via modulating CTGF expression.	
Epigenetics	Inheritable modification in gene expression, Imbalance between HATs AND HDACs	[25,99,100]

reticulum Ca²⁺ ATPase2a (SECRC2a) and well as GLUT4 leading to decreased myocardial functions [59]. Reduced insulin-activated endothelial NO synthase (eNOS) activity and NO production in coronary arteries suppress sarcoplasmic reticulum (SR) Ca²⁺ ion uptake and increase the overload of intracellular Ca²⁺ ions. This results in increased cardiac stiffness and impaired myocardial relaxation in DCM [60].

6.4. Inflammatory cytokines and the innate immune system

Activation of different cells of the innate immune system like neutrophils, mast cells, dendritic cells, macrophages, eosinophils, nuclear transcription factor, NF κ B regulated proinflammatory cytokines like tumor necrosis factor α (TNF α), interleukins (IL) 6 and IL8, monocyte chemotactic protein 1 (MCP1), intercellular adhesion molecule 1 (ICAM1), and vascular cell adhesion molecule 1 (VCAM1) contribute to myocardial oxidative stress, remodeling, fibrosis followed by LV dysfunction [61]. Recently, NACHT, LRR, and PYD domains-containing protein 3 (NLRP-3) were reported to be a novel regulator of inflammation and cellular death via NF κ B and thioredoxin interacting/inhibiting protein (TXNIP) in DCM. Moreover, activated NF κ B also observed to be associated with increased NADP oxidase mediated generation of ROS, peroxynitrite, and superoxide that further increases oxidative stress and decreases ATP synthesis and NO bioavailability and thereby altering cardiac functions in DCM [62].

6.5. Renin angiotensin aldosterone system

In chronic diabetic cases, RAAS gets activated and plays a major role in the pathogenesis of DCM. Evidence suggested the upregulation of proinflammatory angiotensin II receptor 1 (AT2R1) and downregulation of anti-inflammatory AT2R in early DM [63]. Increased plasma aldosterone level and tissue mineralocorticoid receptors (MR) overexpression are linked with insulin resistance, raised BG level, and altered lipid profile (Jia et al., 2017). Studies indicated that inhibition of the aldosterone/MR signaling pathway decreases both morbidity and mortality in diabetic cases with concurrent HF. Enhanced AT2R1 and MR stimulation increases coronary artery endothelial leukocyte/monocyte adhesion, cytokine production, and macrophage infiltration and this results in cardiac remodeling, fibrosis, and diastolic dysfunction in DCM [64].

6.6. Altered substrate utilization and mitochondrial bioenergetics

In the state of persistent hyperglycemia, diabetic myocardium depends almost on the use of FFA for getting ATP and its normal functioning [18]. Many mechanistic approaches are accountable for to shift in substrate utilization. High-fat diet administration showed decreased myocardial GLUT4 levels with alteration in GLUT4 translocation. It causes decreased glycolytic breakdown and glucose oxidation and thus FFA oxidation rates become subsequently increased through the Randle cycle [65,66]. This further activates peroxisome proliferator-activated receptor α (PPAR α) signaling pathways followed by transcriptional

enzyme activation involved in β oxidation. Concurrently, pyruvate dehydrogenase (PDH) was also found to be expressed that further suppresses glucose oxidation [67]. Moreover, evidence showed that myocardial lipotoxicity in DCM cases remains associated with activation of the PPAR- α target gene mCPT1, which controls the mitochondrial FA uptake [68]. This is further supported by other studies that suggest the downregulation of mRNA for GLUT4 and MEF2C in the case of HF of diabetic cases in contrast to failing hearts in patients without diabetes [69]. Similar to myocardial ischemia, altered mitochondrial beta-oxidation of fatty acids has been observed in DCM that even lead to cardiac remodeling and decreased cardiac efficiency [70]. The understanding of mitochondrial bioenergetics, remodeling of the mitochondrial proteome, and reduced respiratory capacity is well studied by using ^{31}P NMR spectroscopy, in both types of DM, independent of CAD [71].

6.7. Advanced glycation end products

Chronic hyperglycemia induces glycation of fibrinogen and albumin resulting in the generation of AGEs that further upregulates the expression of AGEs receptors [72]. Through a series of event, it builds oxidative stress and activates the generation of ROS and inflammatory cytokines, and then promote endothelial and myocardial damage [73]. Increased activation of Janus kinase (JAK), mitogen-activated protein kinase (MAPK) pathway, altered isovolumetric relaxation time and diastolic LV diameter is well evident and reported to be interrelated with myocardial stiffness [74]. AGE production and its cross-linking of collagen molecules in myocardial tissues not only further increases the production of ROS but also it led to the loss of cardiac elasticity due to its fibrosis with subsequent reduction of myocardial compliance [75,76].

6.8. Autophagy

Autophagy is a physiological phenomenon in which long-lived proteins, ribosomes, lipids, and even complete cellular organelles are swallowed up by double-membrane structures, that subsequently targets lysosomes for cellular degradation [77]. Controlled and constitutive autophagy plays a key role in maintaining normal cellular structure and function of cardiac tissue [78]. But variation in this pathway leads to cardiac dysfunction and HF, especially during raised, cellular stress [79]. There is strong evidence that insulin signaling is an important controller of myocardial autophagy [80]. A high fructose diet was reported to be associated with the deposition of autophagosomes as demonstrated by increased levels of the autophagic markers microtubule-associated protein 1A/1B-light chain 3 and nucleoporin p62 [81]. The concurrent raised p62 level further increases the chances of turnover for autophagosomes. Phosphatidylinositol 3-kinase (PI3K) Akt signaling that negatively controls autophagy by inhibiting the mammalian target of rapamycin (MTOR) gets suppressed. Reduced PI3K/Akt signaling starts the possibility of myocardial insulin resistance and thus initiates autophagy [82]. There is a raising consensus that initiation of autophagy can either antagonize disease pathogenesis or contribute to the progression of disease depending on the amplitude of the induction [83]. Therefore, it can be speculated that acute autophagy induction (e.g. ischemic heart) might be beneficial otherwise persistent autophagy induction such as in a diabetic heart could be deleterious [84]. Therefore, autophagy activation in type 1 and type 2 diabetic hearts may either increase or decrease, mediated through PI3K/Akt vs AMPK pathways depending on disease severity.

6.9. MicroRNAs

DCM is related to all-around changes in patterns of gene expression associated with microRNAs (miRNAs). These gene expression regulators are endogenous, noncoding, single-strand RNAs with an average length of 22 nucleotides that remains encoded with small, inverted repeats inside the genome. miRNAs regulate gene expression through the

repression of translation and by promoting the degradation of target mRNAs. Alteration in these miRNA levels exhibits important roles in the pathogenesis of several diseases including DM and associated complications [80]. Recently updated data showed a part of involvement for miRNA-143, miRNA-181, miRNA-103, miRNA-107 and miRNA-802 in the regulation of systemic metabolism of glucose and sensitivity towards insulin. This implicates miRNAs in the pathogenesis of insulin resistance and T2DM [85,86]. A change in myocardial miRNA content is a recognized mechanism that is linked to alteration in cardiac contractility indices. Dysregulation of the miRNA biogenesis in the heart by cardiomyocyte-specific deletion of dicer leads to rapidly progressive dilated cardiomyopathy and HF globally [87]. Recent studies have also connected with the dysregulation of specific miRNAs in DCM [88]. Around 40 % of miRNA-1 of the total myocardial miRNA pool, has been shown to downregulate Pim-1 (proto-oncogene) due to hyperglycemia and its restoration seems to be effective in the prevention of cardiomyocyte apoptosis, ventricular dilatation, and LV failure [89]. Myocardial expression of miRNA-133 has been augmented in DM that further modulates connective tissue content by regulating connective tissue growth factor (CTGF) expression, suggesting its involvement in fibrosis of diabetic hearts [90,91]. The involvement of specific miRNAs in the regulation of systemic metabolism may contribute to identifying the molecular defects associated to DCM [92,93].

6.10. Epigenetics

Epigenetics is an inheritable modification in gene expression patterns, different from alteration in DNA change. Genetic alteration by histone acetylation regulates gene expression. The histone acetylation-mediated genetic alterations are largely regulated through the functional interplay between histone acetyltransferases (HATs), and histone deacetylases (HDACs), which catalyze histone acetylation and deacetylation [94]. Dysregulation of histone acetylation promotes the development of diseases, and HDAC inhibition serves as a therapeutic option to potentially treat a multitude of diseases like cancer and CVDs [95–98]. There are four classes of HDACs identified, depending on sequence identity and domain organization. Out of this, Class II HDACs (HDAC5, HDAC9) can suppress cardiac hypertrophy, while class I HDACs promotes cardiac hypertrophy [99]. Another well-established epigenetic regulator is the methylation of nuclear DNA. DNA methylation takes place mainly on CpG islands (CGIs) in the 5' regulatory regions of many genes. Altogether, epigenetic modifications, which have the potential to influence the expression of the entire genome, may represent an under-investigated mechanism, which may potentially add to the pathogenesis of DCM factors, such as oxidative stress [100].

6.11. Potential chemical compounds:

Considering the duration of clinical studies and cost involved, currently many medications have been proposed to move them forward in the drug discovery to manage DCM. Levosimendan, a potent inodilator, has been reported as a classical inotropes that maintain cardiac mitochondrial energy and thus reducing congestion in acute HF with hypertension. Moreover, with its unique pharmacology it is recommended in wide range of cardiac functional abnormalities related to HF, particularly in undergoing cardiac surgery [101]. A phosphodiesterase type 5 inhibitor (PDE5 inhibitor) reported to reduce reduction in HF-associated hospitalization rate among patients of HF with preserved ejection fraction (HFpEF) and combined pre-capillary and post-capillary pulmonary hypertension [102]. Sildenafil, vardenafil and tadalafil are used clinically an undergoing through intensive investigation for DCM management [103]. Trimetazidine, an anti-ischemic agent that selectively inhibits the long-chain enzyme 3-ketoacyl coenzyme A thiolase, particularly in stressed myocardium and shifts energy utilization from free fatty acid oxidation to glucose oxidation [104,105]. Moreover, reported to improve left ventricular ejection fraction (LVEF), exercise

capacity, and cardiovascular events among patients with DCM [106,107]. Istaxoxime possesses unique luso-inotropic property thus increases SERCA2a pump activity and inhibit Na^+/K^+ ATPase. This improves both myocardial relaxation and contraction including cardiac indexes without severe adverse cardiac events in DCM cases [108–110]. A second line anti-anginal agent for patients with chronic stable angina, ranolazine, mainly acts by inhibiting the late inward sodium current, and improves Ca^{2+} handling and ameliorates impaired myocardial relaxation and diastolic dysfunction in DCM [111–113]. Ranolazine has been reported to decrease left ventricular end-diastolic pressure (LVEDP) and also improved systolic function and reduced cardiovascular events [114,115]. Coenzyme Q10 reported to increase ATP production and cellular energy and thus have cardiovascular protective effects [116]. Improved left ventricular ejection fraction (LVEF) has been reported in a meta-analysis by coenzyme Q10 A supplementation [117,118]. Elamipretide, another mitochondrial antioxidant, stabilizes cardiolipin. Thus, acts as a mitochondrial ROS scavenger and ameliorates left ventricular diastolic dysfunction (LVDF) [119]. A mitochondria-targeted methylglyoxal scavenger, Elamipretide, also stabilizes cardiolipin and scavenges the mitochondrial ROS. Alagebrium, an AGE crosslink breaker, has been also studied extensively. An RCT conducted among both diabetic and non-diabetic patients with diastolic HF showed that it reduces the left ventricular mass, improved diastolic dysfunction, and improved quality of life [120]. However, more preclinical and clinical studies are needed to address safety and efficacy of these drugs.

7. Conclusion

The diabetes-related cardiac disease develops due to various metabolic, structural, and functional alterations. Recent advancements in the understanding of molecular and functional changes within the heart unfold various management opportunities. However, we explored these multifaceted aspects of triggers that are even progressive, for early detection and preventing further related complications that impose a greater healthcare challenge. Therefore, a greater understanding of interrelated pathologies as well as intracellular signaling pathways of diabetic hearts will benefit in tackling and mitigating diabetic cardiomyopathy via novel therapeutic strategies.

Funding

This work was supported by the Dean of Scientific Research, King Khalid University, Saudi Arabia [grant numbers: RA.KKU/30/44].

Declaration of competing interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

Data availability

No data was used for the research described in the article.

Acknowledgements

The authors extend their appreciation to the Deanship of Scientific Research at King Khalid University for funding this work through review article project. (Project under grant number (RA.KKU/30/44).

References

- [1] M.R. Costanzo, Prevention supersedes prediction of worsening heart failure, *JACC Heart Fail.* 11 (2023) 157–160, <https://doi.org/10.1016/j.jchf.2022.11.018>.
- [2] S. Rubler, J. Dlugash, Y.Z. Yuceoglu, T. Kumral, A.W. Branwood, A. Grishman, New type of cardiomyopathy associated with diabetic glomerulosclerosis, *Am. J. Cardiol.* 30 (1972) 595–602, [https://doi.org/10.1016/0002-9149\(72\)90595-4](https://doi.org/10.1016/0002-9149(72)90595-4).
- [3] G. Targher, M. Dauriz, C. Laroche, P.L. Temporelli, M. Hassanein, P.M. Seferovic, J. Drozd, R. Ferrari, S. Anker, A. Coats, G. Filippatos, M.G. Crespo-Leiro, A. Mebazaa, M.F. Piepoli, A.P. Maggioni, L. Tavazzi, ESC-HFA HF long-term registry investigators, in-hospital and 1-year mortality associated with diabetes in patients with acute heart failure: results from the ESC-HFA heart failure long-term registry, *Eur. J. Heart Fail.* 19 (2017) 54–65, <https://doi.org/10.1002/ehf.679>.
- [4] H. Yang, K. Negishi, P. Otahal, T.H. Marwick, Clinical prediction of incident heart failure risk: a systematic review and meta-analysis, *Open Heart.* 2 (2015), e000222, <https://doi.org/10.1136/openhrt-2014-000222>.
- [5] S.S. Jankauskas, U. Kansakar, F. Varzideh, S. Wilson, P. Mone, A. Lombardi, J. Gambardella, G. Santulli, Heart failure in diabetes, *Metabolism.* 125 (2021) 154910, <https://doi.org/10.1016/j.metabol.2021.154910>.
- [6] J.S. Felício, C.C. Koury, C.T. Carvalho, J.F. Abrahão Neto, K.B. Miléo, T. P. Arbage, D.D. Silva, A.F. de Oliveira, A.S. Peixoto, A.B. Figueiredo, A.K. C. Ribeiro Dos Santos, E.S. Yamada, M.T. Zanella, Present insights on cardiomyopathy in diabetic patients, *Curr. Diabetes Rev.* 12 (2016) 384–395, <https://doi.org/10.2174/157339981266150914125029>.
- [7] X. Zhang, H.M. Devlin, B. Smith, G. Imperatore, W. Thomas, F. Lobelo, M.K. Ali, K. Norris, S. Gruss, B. Bardenheier, P. Cho, I. Garcia de Quevedo, U. Mudaliar, C. D. Jones, J.M. Durthaler, J. Saaddine, L.S. Geiss, E.W. Gregg, Effect of lifestyle interventions on cardiovascular risk factors among adults without impaired glucose tolerance or diabetes: a systematic review and meta-analysis, *PLoS One* 12 (2017), e0176436, <https://doi.org/10.1371/journal.pone.0176436>.
- [8] S.S. Mahmood, D. Levy, R.S. Vasan, T.J. Wang, The Framingham Heart Study and the epidemiology of cardiovascular disease: a historical perspective, *Lancet.* 383 (2014) 999–1008, [https://doi.org/10.1016/S0140-6736\(13\)61752-3](https://doi.org/10.1016/S0140-6736(13)61752-3).
- [9] IDF Diabetes Atlas. (n.d.). <https://diabetesatlas.org/ldfawp/resource-files/2021/07/IDFAtlas10thEdition2021.pdf> (accessed March 18, 2023).
- [10] S.S. Virani, A. Alonso, E.J. Benjamin, M.S. Bittencourt, C.W. Callaway, A. P. Carson, A.M. Chamberlain, A.R. Chang, S. Cheng, F.N. Delling, L. Djousse, M.S. V. Elkind, J.F. Ferguson, M. Fornage, S.S. Khan, B.M. Kissela, K.L. Knutson, T. W. Kwan, D.T. Lackland, T.T. Lewis, J.H. Lichtman, C.T. Longenecker, M.S. Loop, P.L. Lutsey, S.S. Martin, K. Matsushita, A.E. Moran, M.E. Mussolino, A.M. Perak, W.D. Rosamond, G.A. Roth, U.K.A. Sampson, G.M. Satou, E.B. Schroeder, S. H. Shah, C.M. Shay, N.L. Spartano, A. Stokes, D.L. Tirschwell, L.B. VanWagner, C. W. Tsao, American Heart Association Council on Epidemiology and Prevention Statistics Committee, Stroke Statistics Subcommittee, Heart disease and stroke statistics-2020 update: a report from the American Heart Association: a report from the American Heart Association, *Circulation* 141 (2020) e139–e596, <https://doi.org/10.1161/CIR.0000000000000757>.
- [11] A. Lorenzo-Almorós, J. Tuñón, M. Orejas, M. Cortés, J. Egido, Ó. Lorenzo, Diagnostic approaches for diabetic cardiomyopathy, *Cardiovasc. Diabetol.* 16 (2017), <https://doi.org/10.1186/s12933-017-0506-x>.
- [12] G. Jia, M.A. Hill, J.R. Sowers, Diabetic cardiomyopathy: an update of mechanisms contributing to this clinical entity, *Circ. Res.* 122 (2018) 624–638, <https://doi.org/10.1161/CIRCRESAHA.117.311586>.
- [13] G. Borghetti, D. von Lewinski, D.M. Eaton, H. Sourij, S.R. Houser, M. Wallner, Diabetic cardiomyopathy: Current and future therapies. Beyond glycemic control, *Front. Physiol.* 9 (2018) 1514, <https://doi.org/10.3389/fphys.2018.01514>.
- [14] S.A. Afanasiev, A.A. Garganeva, E.A. Kuzheleva, A.V. Andriyanova, D. S. Kondratieva, S.V. Popov, Corrigendum to “The impact of type 2 diabetes mellitus on long-term prognosis in patients of different ages with myocardial infarction”, *J. Diabetes Res.* 2019 (2019) 8347891, <https://doi.org/10.1155/2019/8347891>.
- [15] H.-P. Schultheiss, D. Fairweather, A.L.P. Caforio, F. Escher, R.E. Hershberger, S. E. Lipshultz, P.P. Liu, A. Matsumori, A. Mazzanti, J. McMurray, S.G. Priori, Dilated cardiomyopathy, *Nat. Rev. Dis. Primers.* 5 (2019) 32, <https://doi.org/10.1038/s41572-019-0084-1>.
- [16] J.J. Marín-Peñalver, I. Martín-Timón, C. Sevillano-Collantes, F.J. Del Cañizo-Gómez, Update on the treatment of type 2 diabetes mellitus, *World J. Diabetes* 7 (2016) 354–395, <https://doi.org/10.4239/wjcd.v7.i17.354>.
- [17] M. Isfort, S.C.W. Stevens, S. Schaffer, C.J. Jong, L.E. Wold, Metabolic dysfunction in diabetic cardiomyopathy, *Heart Fail. Rev.* 19 (2014) 35–48, <https://doi.org/10.1007/s10741-013-9377-8>.
- [18] J. Fuentes-Antrás, B. Picatoste, E. Ramírez, J. Egido, J. Tuñón, Ó. Lorenzo, Targeting metabolic disturbance in the diabetic heart, *Cardiovasc. Diabetol.* 14 (2015) 17, <https://doi.org/10.1186/s12933-015-0173-8>.
- [19] Y.K. Bando, T. Murohara, Diabetes-related heart failure: - does diabetic cardiomyopathy exist? *Circ. J.* 78 (2014) 576–583, <https://doi.org/10.1253/circj.13-1564>.
- [20] D.Z. Mytas, P.N. Stogiannos, M.N. Zairis, S.G. Foussas, V.N. Pyrgakis, I. A. Kyriazis, Diabetic myocardial disease: pathophysiology, early diagnosis and therapeutic options, *J. Diabetes Complicat.* 23 (2009) 273–282, <https://doi.org/10.1016/j.jdiacomp.2007.12.005>.
- [21] T. Miki, S. Yuda, H. Kouzu, T. Miura, Diabetic cardiomyopathy: pathophysiology and clinical features, *Heart Fail. Rev.* 18 (2013) 149–166, <https://doi.org/10.1007/s10741-012-9313-3>.
- [22] B.R. Goyal, A.A. Mehta, Diabetic cardiomyopathy: pathophysiological mechanisms and cardiac dysfunction: pathophysiological mechanisms and cardiac dysfunction, *Hum. Exp. Toxicol.* 32 (2013) 571–590, <https://doi.org/10.1177/0960327112450885>.

- [23] V.R. Taqueti, M.F. Di Carli, Coronary microvascular disease pathogenic mechanisms and therapeutic options: JACC state-of-the-art review, *J. Am. Coll. Cardiol.* 72 (2018) 2625–2641, <https://doi.org/10.1016/j.jacc.2018.09.042>.
- [24] F. Vancheri, G. Longo, S. Vancheri, M. Henein, Coronary microvascular dysfunction, *J. Clin. Med.* 9 (2020) 2880, <https://doi.org/10.3390/jcm9092880>.
- [25] J.D. Sara, R. Taher, N. Kolluri, A. Vella, L.O. Lerman, A. Lerman, Coronary microvascular dysfunction is associated with poor glycemic control amongst female diabetics with chest pain and non-obstructive coronary artery disease, *Cardiovasc. Diabetol.* 18 (2019) 22, <https://doi.org/10.1186/s12933-019-0833-1>.
- [26] R.T. Cowling, D. Kupsky, A.M. Kahn, L.B. Daniels, B.H. Greenberg, Mechanisms of cardiac collagen deposition in experimental models and human disease, *Transl. Res.* 209 (2019) 138–155, <https://doi.org/10.1016/j.trsl.2019.03.004>.
- [27] H. Tsutsui, S. Matsushima, S. Kinugawa, T. Ide, N. Inoue, Y. Ohta, T. Yokota, S. Hamaguchi, K. Sunagawa, Angiotensin II type 1 receptor blocker attenuates myocardial remodeling and preserves diastolic function in diabetic heart, *Hypertens. Res.* 30 (2007) 439–449, <https://doi.org/10.1291/hyres.30.439>.
- [28] C.R. Ban, S.M. Twigg, Fibrosis in diabetes complications: pathogenic mechanisms and circulating and urinary markers, *Vasc. Health Risk Manag.* 4 (2008) 575–596, <https://doi.org/10.2147/vhrm.s1991>.
- [29] Y. Chen, Y. Hua, X. Li, I.M. Arslan, W. Zhang, G. Meng, Distinct types of cell death and the implication in diabetic cardiomyopathy, *Front. Pharmacol.* 11 (2020) 42, <https://doi.org/10.3389/fphar.2020.00042>.
- [30] E. Shen, Y. Li, Y. Li, L. Shan, H. Zhu, Q. Feng, J.M.O. Arnold, T. Peng, Rac1 is required for cardiomyocyte apoptosis during hyperglycemia, *Diabetes* 58 (2009) 2386–2395, <https://doi.org/10.2337/db08-0617>.
- [31] S. Bernardi, A. Michelli, G. Zuolo, R. Candido, B. Fabris, Update on RAAS modulation for the treatment of diabetic cardiovascular disease, *J. Diabetes Res.* 2016 (2016) 8917578, <https://doi.org/10.1155/2016/8917578>.
- [32] A.I. Vinik, C. Casellini, H.K. Parson, S.R. Colberg, M.-L. Nevoret, Cardiac autonomic neuropathy in diabetes: a predictor of cardiometabolic events, *Front. Neurosci.* 12 (2018) 591, <https://doi.org/10.3389/fnins.2018.00591>.
- [33] J.M.G. Fernandes, B. de Oliveira Romão, I.R. Rivera, M.A. Mendonça, F. de A. Costa, M. de S. Lira Handro, O. Campos, Á.A.V. De Paola, V.A. Moisés, Clinical value of myocardial performance index in patients with isolated diastolic dysfunction, *Cardiovasc. Ultrasound* 17 (2019) 17, <https://doi.org/10.1186/s12947-019-0167-x>.
- [34] M. Galderisi, K.M. Anderson, P.W. Wilson, D. Levy, Echocardiographic evidence for the existence of a distinct diabetic cardiomyopathy (the Framingham Heart Study), *Am. J. Cardiol.* 68 (1991) 85–89, [https://doi.org/10.1016/0002-9149\(91\)90716-x](https://doi.org/10.1016/0002-9149(91)90716-x).
- [35] K. Eguchi, B. Boden-Albala, Z. Jin, T. Rundek, R.L. Sacco, S. Homma, M.R. Di Tullio, Association between diabetes mellitus and left ventricular hypertrophy in a multiethnic population, *Am. J. Cardiol.* 101 (2008) 1787–1791, <https://doi.org/10.1016/j.amjcard.2008.02.082>.
- [36] P. Perkkatannapatt, R.B. D'Agostino Jr., K.M. Link, E. Shahar, J.A. Lima, D. A. Blumke, S. Sinha, D.M. Herrington, W.G. Hundley, Location of arterial stiffening differs in those with impaired fasting glucose versus diabetes: implications for left ventricular hypertrophy from the multi-ethnic study of atherosclerosis, *Diabetes* 58 (2009) 946–953, <https://doi.org/10.2337/db08-1192>.
- [37] Z.Y. Fang, R. Schull-Meade, R. Leano, P.M. Mottram, J.B. Prins, T.H. Marwick, Screening for heart disease in diabetic subjects, *Am. Heart J.* 149 (2005) 349–354, <https://doi.org/10.1016/j.ahj.2004.06.021>.
- [38] B.A. Brooks, B. Franjic, C.R. Ban, K. Swaraj, D.K. Yue, D.S. Celermajer, S. M. Twigg, Diastolic dysfunction and abnormalities of the microcirculation in type 2 diabetes, *Diabetes Obes. Metab.* 10 (2008) 739–746, <https://doi.org/10.1111/j.1463-1326.2007.00803.x>.
- [39] R.W. van der Meer, L.J. Rijzewijk, H.W.A.M. de Jong, H.J. Lamb, M. Lubberink, J. A. Romijn, J.J. Bax, A. de Roos, O. Kamp, W.J. Paulus, R.J. Heine, A. A. Lammertsma, J.W.A. Smit, M. Diamant, Pioglitazone improves cardiac function and alters myocardial substrate metabolism without affecting cardiac triglyceride accumulation and high-energy phosphate metabolism in patients with well-controlled type 2 diabetes mellitus, *Circulation* 119 (2009) 2069–2077, <https://doi.org/10.1161/CIRCULATIONAHA.108.803916>.
- [40] P.H. Waddingham, S. Bhattacharyya, J. Van Zalen, G. Lloyd, Contractile reserve as a predictor of prognosis in patients with non-ischaemic systolic heart failure and dilated cardiomyopathy: a systematic review and meta-analysis, *Echo Res. Pract.* 5 (2018) 1–9, <https://doi.org/10.1530/ERP-17-0054>.
- [41] D. Joshi, R. Gupta, A. Dubey, A. Shiwalkar, P. Pathak, R.C. Gupta, V. Chauthaiwale, C. Dutt, TRC4186, a novel AGE-breaker, improves diabetic cardiomyopathy and nephropathy in Ob-ZSF1 model of type 2 diabetes, *J. Cardiovasc. Pharmacol.* 54 (2009) 72–81, <https://doi.org/10.1097/FJC.0b013e3181ac3a34>.
- [42] V. Lamounier-Zepter, C. Look, J. Alvarez, T. Christ, U. Ravens, W.-H. Schunck, M. Ehrhart-Bornstein, S.R. Bornstein, I. Morano, Adipocyte fatty acid-binding protein suppresses cardiomyocyte contraction: a new link between obesity and heart disease: a new link between obesity and heart disease, *Circ. Res.* 105 (2009) 326–334, <https://doi.org/10.1161/CIRCRESAHA.109.200501>.
- [43] S.A. Cook, A. Varela-Carver, M. Mongillo, C. Kleinert, M.T. Khan, L. Leccisotti, N. Strickland, T. Matsui, S. Das, A. Rosenzweig, P. Punjabi, P.G. Camici, Abnormal myocardial insulin signalling in type 2 diabetes and left-ventricular dysfunction, *Eur. Heart J.* 31 (2010) 100–111, <https://doi.org/10.1093/eurheartj/ehp396>.
- [44] F. Dong, X. Zhang, X. Yang, L.B. Esberg, H. Yang, Z. Zhang, B. Culver, J. Ren, Impaired cardiac contractile function in ventricular myocytes from leptin-deficient ob/ob obese mice, *J. Endocrinol.* 188 (2006) 25–36, <https://doi.org/10.1677/joe.1.06241>.
- [45] S. Sharma, J.V. Adrogué, L. Gofman, I. Uray, J. Lemm, K. Youker, G.P. Noon, O. H. Frazier, H. Taegtmeyer, Intramyocardial lipid accumulation in the failing human heart resembles the lipotoxic rat heart, *FASEB J.* 18 (2004) 1692–1700, <https://doi.org/10.1096/fj.04-2263com>.
- [46] A.C. Sletten, L.R. Peterson, J.E. Schaffer, Manifestations and mechanisms of myocardial lipotoxicity in obesity, *J. Intern. Med.* 284 (2018) 478–491, <https://doi.org/10.1111/joim.12728>.
- [47] T.J. Regan, M.M. Lyons, S.S. Ahmed, G.E. Levinson, H.A. Oldewurtel, M. R. Ahmad, B. Haider, Evidence for cardiomyopathy in familial diabetes mellitus, *J. Clin. Invest.* 60 (1977) 884–899, <https://doi.org/10.1172/JCI108843>.
- [48] J.M. McGavock, I. Lingvay, I. Zib, T. Tillery, N. Salas, R. Unger, B.D. Levine, P. Raskin, R.G. Victor, L.S. Szczepaniak, Cardiac steatosis in diabetes mellitus: a 1H-magnetic resonance spectroscopy study: a 1 H-magnetic resonance spectroscopy study, *Circulation* 116 (2007) 1170–1175, <https://doi.org/10.1161/CIRCULATIONAHA.106.645614>.
- [49] T.-S. Park, Y. Hu, H.-L. Noh, K. Drosatos, K. Okajima, J. Buchanan, J. Tuinei, S. Homma, X.-C. Jiang, E.D. Abel, L.J. Goldberg, Ceramide is a cardiotoxin in lipotoxic cardiomyopathy, *J. Lipid Res.* 49 (2008) 2101–2112, <https://doi.org/10.1194/jlr.M800147-JLR200>.
- [50] N.M. Borradaile, X. Han, J.D. Harp, S.E. Gale, D.S. Ory, J.E. Schaffer, Disruption of endoplasmic reticulum structure and integrity in lipotoxic cell death, *J. Lipid Res.* 47 (2006) 2726–2737, <https://doi.org/10.1194/jlr.M600299-JLR200>.
- [51] R.T. Brookheart, C.I. Michel, L.L. Listenberger, D.S. Ory, J.E. Schaffer, The non-coding RNA gadd7 is a regulator of lipid-induced oxidative and endoplasmic reticulum stress, *J. Biol. Chem.* 284 (2009) 7446–7454, <https://doi.org/10.1074/jbc.M806209200>.
- [52] N.S. Dhalla, A.K. Shah, P.S. Tappia, Role of oxidative stress in metabolic and subcellular abnormalities in diabetic cardiomyopathy, *Int. J. Mol. Sci.* 21 (2020) 2413, <https://doi.org/10.3390/ijms21072413>.
- [53] S. Serpillon, B.C. Floyd, R.S. Gupta, S. George, M. Kozicky, V. Neito, F. Recchia, W. Stanley, M.S. Wolin, S.A. Gupta, Superoxide production by NAD(P)H oxidase and mitochondria is increased in genetically obese and hyperglycemic rat heart and aorta before the development of cardiac dysfunction. The role of glucose-6-phosphate dehydrogenase-derived NADPH, *Am. J. Physiol. Heart Circ. Physiol.* 297 (2009) H153–H162, <https://doi.org/10.1152/ajpheart.01142.2008>.
- [54] R. Alhayaza, E. Haque, C. Karbasiafshar, F.W. Sellke, M.R. Abid, The relationship between reactive oxygen species and endothelial cell metabolism, *Front. Chem.* 8 (2020), <https://doi.org/10.3389/fchem.2020.592688>.
- [55] S. Freitas-Rodríguez, A.R. Folgueras, C. López-Otín, The role of matrix metalloproteinases in aging: tissue remodeling and beyond, *Biochim. Biophys. Acta, Mol. Cell Res.* 1864 (2017) 2015–2025, <https://doi.org/10.1016/j.bbamcr.2017.05.007>.
- [56] J.K. Seok, H.C. Kang, Y.-Y. Cho, H.S. Lee, J.Y. Lee, Therapeutic regulation of the NLRP3 inflammasome in chronic inflammatory diseases, *Arch. Pharm. Res.* 44 (2021) 16–35, <https://doi.org/10.1007/s12272-021-01307-9>.
- [57] G. Santulli, D.R. Lewis, A.R. Marks, Physiology and pathophysiology of excitation-contraction coupling: the functional role of ryanodine receptor, *J. Muscle Res. Cell Motil.* 38 (2017) 37–45, <https://doi.org/10.1007/s10974-017-9470-z>.
- [58] R. Nikolaenko, E. Bovo, A.V. Zima, Redox dependent modifications of ryanodine receptor: basic mechanisms and implications in heart diseases, *Front. Physiol.* 9 (2018) 1775, <https://doi.org/10.3389/fphys.2018.01775>.
- [59] M. Zamora, J.A. Villena, Contribution of impaired insulin signaling to the pathogenesis of diabetic cardiomyopathy, *Int. J. Mol. Sci.* 20 (2019) 2833, <https://doi.org/10.3390/ijms20112833>.
- [60] R. Muniyappa, M. Montagnani, K.K. Koh, M.J. Quon, Cardiovascular actions of insulin, *Endocr. Rev.* 28 (2007) 463–491, <https://doi.org/10.1210/er.2007-0006>.
- [61] A. Zimmer, A.K. Bagchi, K. Vinayak, A. Bello-Klein, P.K. Singal, Innate immune response in the pathogenesis of heart failure in survivors of myocardial infarction, *Am. J. Physiol. Heart Circ. Physiol.* 316 (2019) H435–H445, <https://doi.org/10.1152/ajpheart.00597.2018>.
- [62] B. Luo, F. Huang, Y. Liu, Y. Liang, Z. Wei, H. Ke, Z. Zeng, W. Huang, Y. He, NLRP3 inflammasome as a molecular marker in diabetic cardiomyopathy, *Front. Physiol.* 8 (2017) 519, <https://doi.org/10.3389/fphys.2017.00519>.
- [63] S.J. Forrester, G.W. Booz, C.D. Sigmund, T.M. Coffman, T. Kawai, V. Rizzo, R. Scalia, S. Eguchi, Angiotensin II signal transduction: an update on mechanisms of physiology and pathophysiology, *Physiol. Rev.* 98 (2018) 1627–1738, <https://doi.org/10.1152/physrev.00038.2017>.
- [64] G. Jia, J. Habibi, V.G. DeMarco, L.A. Martinez-Lemus, L. Ma, A.T. Whaley-Connell, A.R. Aroor, T.L. Domeier, Y. Zhu, G.A. Meininger, K.B. Mueller, I. Z. Jaffe, J.R. Sowers, Endothelial mineralocorticoid receptor deletion prevents diet-induced cardiac diastolic dysfunction in females, *Hypertension* 66 (2015) 1159–1167, <https://doi.org/10.1161/HYPERTENSIONAHA.115.06015>.
- [65] Y. Wang, L. Wen, S. Zhou, Y. Zhang, X.-H. Wang, Y.-Y. He, A. Davie, S. Broadbent, Effects of four weeks intermittent hypoxia intervention on glucose homeostasis, insulin sensitivity, GLUT4 translocation, insulin receptor phosphorylation, and Akt activity in skeletal muscle of obese mice with type 2 diabetes, *PLoS One* 13 (2018), e0203551, <https://doi.org/10.1371/journal.pone.0203551>.
- [66] P.J. Randle, P.B. Garland, C.N. Hales, E.A. Newsholme, The glucose fatty-acid cycle its role in insulin sensitivity and the metabolic disturbances of diabetes mellitus, *Lancet* 281 (1963) 785–789, [https://doi.org/10.1016/s0140-6736\(63\)91500-9](https://doi.org/10.1016/s0140-6736(63)91500-9).
- [67] J.J. Wright, J. Kim, J. Buchanan, S. Boudina, S. Sena, K. Bakirtzi, O. Ilkun, H. A. Theobald, R.C. Cooksey, K.V. Kandror, E.D. Abel, Mechanisms for increased

- myocardial fatty acid utilization following short-term high-fat feeding, *Cardiovasc. Res.* 82 (2009) 351–360, <https://doi.org/10.1093/cvr/cvp017>.
- [68] L. Wang, Y. Cai, L. Jian, C.W. Cheung, L. Zhang, Z. Xia, Impact of peroxisome proliferator-activated receptor- α on diabetic cardiomyopathy, *Cardiovasc. Diabetol.* 20 (2021) 2, <https://doi.org/10.1186/s12933-020-01188-0>.
- [69] P. Razeghi, M.E. Young, T.C. Cockrill, O.H. Frazier, H. Taegtmeier, Downregulation of myocardial myocyte enhancer factor 2C and myocyte enhancer factor 2C-regulated gene expression in diabetic patients with nonischemic heart failure, *Circulation*. 106 (2002) 407–411, <https://doi.org/10.1161/01.cir.0000026392.80723.dc>.
- [70] Q.G. Karwi, G.M. Uddin, K.L. Ho, G.D. Lopaschuk, Loss of metabolic flexibility in the failing heart, *Front. Cardiovasc. Med.* 5 (2018), <https://doi.org/10.3389/fcvm.2018.00068>.
- [71] D. Abdurrahim, J.J. Prompers, Evaluation of cardiac energetics by non-invasive ³¹P magnetic resonance spectroscopy, *Biochim. Biophys. Acta Mol. basis Dis.* 1864 (2018) 1939–1948, <https://doi.org/10.1016/j.bbdis.2017.11.013>.
- [72] L. Egaña-Gorroño, R. López-Díez, G. Yepuri, L.S. Ramirez, S. Reverdatto, P. F. Gugger, A. Shekhtman, R. Ramasamy, A.M. Schmidt, Receptor for advanced glycation end products (RAGE) and mechanisms and therapeutic opportunities in diabetes and cardiovascular disease: insights from human subjects and animal models, *Front. Cardiovasc. Med.* 7 (2020) 37, <https://doi.org/10.3389/fcvm.2020.00037>.
- [73] S.L. Fishman, H. Sonmez, C. Basman, V. Singh, L. Poretsky, The role of advanced glycation end-products in the development of coronary artery disease in patients with and without diabetes mellitus: a review, *Mol. Med.* 24 (2018) 59, <https://doi.org/10.1186/s10020-018-0060-3>.
- [74] N.N. Mehta, Potential cardiovascular implications of Janus kinase inhibitors in immune mediated diseases, *Cardiovasc. Res.* 114 (2018) e81–e83, <https://doi.org/10.1093/cvr/cvy160>.
- [75] M. Kosmopoulos, D. Drekolias, P.D. Zavras, C. Piperi, A.G. Papavassiliou, Impact of advanced glycation end products (AGEs) signaling in coronary artery disease, *Biochim. Biophys. Acta Mol. basis Dis.* 1865 (2019) 611–619, <https://doi.org/10.1016/j.bbdis.2019.01.006>.
- [76] R. D'Oria, R. Schipani, A. Leonardini, A. Natalicchio, S. Perrini, A. Cignarelli, L. Laviola, F. Giorgino, The role of oxidative stress in cardiac disease: from physiological response to injury factor, *Oxidative Med. Cell. Longev.* 2020 (2020) 5732956, <https://doi.org/10.1155/2020/5732956>.
- [77] P. Ravanan, I.F. Srikumar, P. Talwar, Autophagy: the spotlight for cellular stress responses, *Life Sci.* 188 (2017) 53–67, <https://doi.org/10.1016/j.lfs.2017.08.029>.
- [78] L. Galluzzi, F. Pietrocola, B. Levine, G. Kroemer, Metabolic control of autophagy, *Cell.* 159 (2014) 1263–1276, <https://doi.org/10.1016/j.cell.2014.11.006>.
- [79] S. Sciarretta, Y. Maejima, D. Zablocki, J. Sadoshima, The role of autophagy in the heart, *Annu. Rev. Physiol.* 80 (2018) 1–26, <https://doi.org/10.1146/annurev-physiol-021317-121427>.
- [80] R. Guo, S. Nair, Role of microRNA in diabetic cardiomyopathy: from mechanism to intervention, *Biochim. Biophys. Acta Mol. basis Dis.* 1863 (2017) 2070–2077, <https://doi.org/10.1016/j.bbdis.2017.03.013>.
- [81] M.W. Baeken, K. Weckmann, P. Diefenthaler, J. Schulte, K. Yusifli, B. Moosmann, C. Behl, P. Hajieva, Novel insights into the cellular localization and regulation of the autophagosomal proteins LC3A, LC3B and LC3C, *Cells.* 9 (2020) 2315, <https://doi.org/10.3390/cells9102315>.
- [82] J. Zhao, J.J. Brault, A. Schild, P. Cao, M. Sandri, S. Schiaffino, S.H. Lecker, A. L. Goldberg, FoxO3 coordinately activates protein degradation by the autophagic/lysosomal and proteasomal pathways in atrophying muscle cells, *Cell Metab.* 6 (2007) 472–483, <https://doi.org/10.1016/j.cmet.2007.11.004>.
- [83] G.G. Schiattarella, J.A. Hill, Therapeutic targeting of autophagy in cardiovascular disease, *J. Mol. Cell. Cardiol.* 95 (2016) 86–93, <https://doi.org/10.1016/j.yjmcc.2015.11.019>.
- [84] Z. Xie, K. Lau, B. Eby, P. Lozano, C. He, B. Pennington, H. Li, S. Rath, Y. Dong, R. Tian, D. Kem, M.-H. Zou, Improvement of cardiac functions by chronic metformin treatment is associated with enhanced cardiac autophagy in diabetic OVE26 mice, *Diabetes.* 60 (2011) 1770–1778, <https://doi.org/10.2337/db10-0351>.
- [85] S.D. Jordan, M. Krüger, D.M. Willmes, N. Redemann, F.T. Wunderlich, H. S. Brönneke, C. Merkwirth, H. Kashkar, V.M. Olkkonen, T. Böttger, T. Braun, J. Seibler, J.C. Brüning, Obesity-induced overexpression of miRNA-143 inhibits insulin-stimulated AKT activation and impairs glucose metabolism, *Nat. Cell Biol.* 13 (2011) 434–446, <https://doi.org/10.1038/ncb2211>.
- [86] M. Trajkovski, J. Hausser, J. Soutschek, B. Bhat, A. Akin, M. Zavolan, M.H. Heim, M. Stoffel, MicroRNAs 103 and 107 regulate insulin sensitivity, *Nature.* 474 (2011) 649–653, <https://doi.org/10.1038/nature10112>.
- [87] J.-F. Chen, E.P. Murchison, R. Tang, T.E. Callis, M. Tatsuguchi, Z. Deng, M. Rojas, S.M. Hammond, M.D. Schneider, C.H. Selzman, G. Meissner, C. Patterson, G. J. Hannon, D.-Z. Wang, Targeted deletion of dicer in the heart leads to dilated cardiomyopathy and heart failure, *Proc. Natl. Acad. Sci. U. S. A.* 105 (2008) 2111–2116, <https://doi.org/10.1073/pnas.0710228105>.
- [88] R.V. Kartha, S. Subramanian, MicroRNAs in cardiovascular diseases: biology and potential clinical applications, *J. Cardiovasc. Transl. Res.* 3 (2010) 256–270, <https://doi.org/10.1007/s12265-010-9172-z>.
- [89] R. Katare, A. Caporali, L. Zentilin, E. Avolio, G. Sala-Newby, A. Oikawa, D. Cesselli, A.P. Beltrami, M. Giacca, C. Emanueli, P. Madeddu, Intravenous gene therapy with PIM-1 via a cardiotropic viral vector halts the progression of diabetic cardiomyopathy through promotion of pro-survival signaling, *Circ. Res.* 108 (2011) 1238–1251, <https://doi.org/10.1161/CIRCRESAHA.110.239111>.
- [90] J. Xiao, X. Luo, H. Lin, Y. Zhang, Y. Lu, N. Wang, Y. Zhang, B. Yang, Z. Wang, MicroRNA miR-133 represses HERG K⁺ channel expression contributing to QT prolongation in diabetic hearts, *J. Biol. Chem.* 286 (2011) 28656, <https://doi.org/10.1074/jbc.a111.700015>.
- [91] R.F. Duisters, A.J. Tijssen, B. Schroen, J.J. Leenders, V. Lentink, I. van der Made, V. Herias, R.E. van Leeuwen, M.W. Schellings, P. Barenbrug, J.G. Maessen, S. Heymans, Y.M. Pinto, E.E. Creemers, miR-133 and miR-30 regulate connective tissue growth factor: implications for a role of microRNAs in myocardial matrix remodeling: implications for a role of MicroRNAs in myocardial matrix remodeling, *Circ. Res.* 104 (2009) 170–178, 6p following 178, <https://doi.org/10.1161/CIRCRESAHA.108.182535>.
- [92] R.C. Friedman, K.K.-H. Farh, C.B. Burge, D.P. Bartel, Most mammalian mRNAs are conserved targets of microRNAs, *Genome Res.* 19 (2009) 92–105, <https://doi.org/10.1101/gr.082701.108>.
- [93] S. Shantikumar, A. Caporali, C. Emanueli, Role of microRNAs in diabetes and its cardiovascular complications, *Cardiovasc. Res.* 93 (2012) 583–593, <https://doi.org/10.1093/cvr/cvr300>.
- [94] S.P. Barros, S. Offenbacher, Epigenetics: connecting environment and genotype to phenotype and disease, *J. Dent. Res.* 88 (2009) 400–408, <https://doi.org/10.1177/0022034509335868>.
- [95] D.-M. Chuang, Y. Leng, Z. Marinova, H.-J. Kim, C.-T. Chiu, Multiple roles of HDAC inhibition in neurodegenerative conditions, *Trends Neurosci.* 32 (2009) 591–601, <https://doi.org/10.1016/j.tins.2009.06.002>.
- [96] O. Khan, N.B. La Thangue, HDAC inhibitors in cancer biology: emerging mechanisms and clinical applications, *Immunol. Cell Biol.* 90 (2012) 85–94, <https://doi.org/10.1038/icb.2011.100>.
- [97] T.A. McKinsey, Isoform-selective HDAC inhibitors: closing in on translational medicine for the heart, *J. Mol. Cell. Cardiol.* 51 (2011) 491–496, <https://doi.org/10.1016/j.yjmcc.2010.11.009>.
- [98] M.E. Pepin, A.R. Wende, Epigenetics in the development of diabetic cardiomyopathy, *Epigenomics.* 11 (2019) 469–472, <https://doi.org/10.2217/epi-2019-0027>.
- [99] S.-Y. Park, J.-S. Kim, A short guide to histone deacetylases including recent progress on class II enzymes, *Exp. Mol. Med.* 52 (2020) 204–212, <https://doi.org/10.1038/s12276-020-0382-4>.
- [100] L.M. Villeneuve, R. Natarajan, The role of epigenetics in the pathology of diabetic complications, *Am. J. Physiol. Ren. Physiol.* 299 (2010) F14–F25, <https://doi.org/10.1152/ajprenal.00200.2010>.
- [101] M.S. Akhtar, M.Q. Hassan, A. Siddiqui, S.S. Alavudeen, O. Afzal, A.S.A. Altamimi, S.O. Rahman, M. Khurana, M.J. Ahsan, A.K. Sharma, F. Tabassum, Levosimendan: mechanistic insight and its diverse future aspects in cardiac care, *Acta Cardiol.* 78 (2023) 170–187, <https://doi.org/10.1080/00015385.2022.2115761>.
- [102] M. Guazzi, Clinical use of phosphodiesterase-5 inhibitors in chronic heart failure, *Circ. Heart Fail.* 1 (2008) 272–280, <https://doi.org/10.1161/CIRCHEARTFAILURE.108.802116>.
- [103] C.D. Boyle, R. Xu, T. Asberom, S. Chackalamannil, J.W. Clader, W.J. Greenlee, H. Guzik, Y. Hu, Z. Hu, C.M. Lankin, D.A. Pissarnitski, A.W. Stamford, Y. Wang, J. Skell, S. Kuroski, S. Vemulapalli, J. Palamanda, M. Chintala, P. Wu, J. Myers, P. Wang, Optimization of purine based PDE1/PDE5 inhibitors to a potent and selective PDE5 inhibitor for the treatment of male ED, *Bioorg. Med. Chem. Lett.* 15 (2005) 2365–2369, <https://doi.org/10.1016/j.bmcl.2005.02.083>.
- [104] H. Shu, Y. Peng, W. Hang, N. Zhou, D.W. Wang, Trimetazidine in heart failure, *Front. Pharmacol.* 11 (2021), <https://doi.org/10.3389/fphar.2020.569132>.
- [105] M. Marzilli, D. Vinereanu, G. Lopaschuk, Y. Chen, J.J. Dalal, N. Danchin, E. Etriby, R. Ferrari, L.H. Gowdak, Y. Lopatin, D. Milicic, A. Parkhomenko, F. Pinto, P. Ponikowski, P. Seferovic, G.M.C. Rosano, Trimetazidine in cardiovascular medicine, *Int. J. Cardiol.* 293 (2019) 39–44, <https://doi.org/10.1016/j.ijcard.2019.05.063>.
- [106] D. Gao, N. Ning, X. Niu, G. Hao, Z. Meng, Trimetazidine: a meta-analysis of randomised controlled trials in heart failure, *Heart.* 97 (2011) 278–286, <https://doi.org/10.1136/hrt.2010.208751>.
- [107] G. Fragasso, A. Salerno, G. Lattuada, A. Cuko, G. Calori, A. Scollo, F. Ragogna, F. Arioli, G. Bassanelli, R. Spoladore, L. Luzi, A. Margonato, G. Perseghini, Effect of partial inhibition of fatty acid oxidation by trimetazidine on whole body energy metabolism in patients with chronic heart failure, *Heart.* 97 (2011) 1495–1500, <https://doi.org/10.1136/hrt.2011.226332>.
- [108] H. Khan, M. Metra, J.E.A. Blair, M. Vogel, M.E. Harinstein, G.S. Filippatos, H. N. Sabbah, H. Porchet, G. Valentini, M. Gheorghiad, Istaroxime, a first in class new chemical entity exhibiting SERCA-2 activation and Na-K-ATPase inhibition: a new promising treatment for acute heart failure syndromes? *Heart Fail. Rev.* 14 (2009) 277–287, <https://doi.org/10.1007/s10741-009-9136-z>.
- [109] S.J. Shah, J.E.A. Blair, G.S. Filippatos, C. Macarie, W. Ruzyllo, J. Korewicki, S. I. Bubenek-Turconi, M. Ceracchi, M. Bianchetti, P. Carminati, D. Kremastinos, J. Grzybowski, G. Valentini, H.N. Sabbah, M. Gheorghiad, HORIZON-HF Investigators, Effects of istaroxime on diastolic stiffness in acute heart failure syndromes: results from the hemodynamic, echocardiographic, and neurohormonal effects of istaroxime, a novel intravenous inotropic and lusitropic agent: a randomized controlled trial in patients hospitalized with heart failure (HORIZON-HF) trial, *Am. Heart J.* 157 (2009) 1035–1041, <https://doi.org/10.1016/j.ahj.2009.03.007>.
- [110] V. Carubelli, Y. Zhang, M. Metra, C. Lombardi, G.M. Felker, G. Filippatos, C. M. O'Connor, J.R. Teerlink, P. Simmons, R. Segal, G. Malfatto, M.T. La Rovere, D. Li, X. Han, Z. Yuan, Y. Yao, B. Li, L.F. Lau, G. Bianchi, J. Zhang, the Istaroxime ADHF Trial Group, Treatment with 24 hour istaroxime infusion in patients hospitalised for acute heart failure: a randomised, placebo-controlled trial, *Eur. J. Heart Fail.* 22 (2020) 1684–1693, <https://doi.org/10.1002/ehf.1743>.

- [111] S.D. Fihn, J.M. Gardin, J. Abrams, K. Berra, J.C. Blankenship, A.P. Dallas, P. S. Douglas, J.M. Foody, T.C. Gerber, A.L. Hinderliter, S.B. King III, P.D. Kligfield, H.M. Krumholz, R.Y.K. Kwong, M.J. Lim, J.A. Linderbaum, M.J. Mack, M. A. Munger, R.L. Prager, J.F. Sabik, L.J. Shaw, J.D. Sikkema, C.R. Smith Jr., S. C. Smith Jr., J.A. Spertus, S.V. Williams, 2012 ACCF/AHA/ACP/AATS/PCNA/SCAI/STS guideline for the diagnosis and management of patients with stable ischemic heart disease: executive summary, *J. Am. Coll. Cardiol.* 60 (2012) 2564–2603, <https://doi.org/10.1016/j.jacc.2012.07.012>.
- [112] Task Force Members, G. Montalescot, U. Sechtem, S. Achenbach, F. Andreotti, C. Arden, A. Budaj, R. Bugiardini, F. Crea, T. Cuisset, C. Di Mario, J.R. Ferreira, B. Gersh, A.K. Gitt, J.-S. Hulot, N. Marx, L.H. Opie, M. Pfisterer, E. Prescott, F. Ruschitzka, M. Sabaté, R. Senior, D.P. Taggart, E.E. van der Wall, C.J.M. Vrints, ESC Committee for Practice Guidelines, J.L. Zamorano, S. Achenbach, H. Baumgartner, J.J. Bax, H. Bueno, V. Dean, C. Deaton, C. Erol, R. Fagard, R. Ferrari, D. Hasdai, A.W. Hoes, P. Kirchhof, J. Knuuti, P. Kolh, P. Lancellotti, A. Linhart, P. Nihoyannopoulos, M.F. Piepoli, P. Ponikowski, P.A. Sirnes, J. L. Tamargo, M. Tendera, A. Torbicki, W. Wijns, S. Windecker, Document Reviewers, J. Knuuti, M. Valgimigli, H. Bueno, M.J. Claeys, N. Donner-Banzhoff, C. Erol, H. Frank, C. Funck-Brentano, O. Gaemperli, J.R. Gonzalez-Juanatey, M. Hamilos, D. Hasdai, S. Husted, S.K. James, K. Kervinen, P. Kolh, S. D. Kristensen, P. Lancellotti, A.P. Maggioni, M.F. Piepoli, A.R. Pries, F. Romeo, L. Rydén, M.L. Simoons, P.A. Sirnes, P.G. Steg, A. Timmis, W. Wijns, S. Windecker, A. Yildirim, J.L. Zamorano, 2013 ESC guidelines on the management of stable coronary artery disease: the task force on the management of stable coronary artery disease of the European Society of Cardiology: the task force on the management of stable coronary artery disease of the European Society of Cardiology, *Eur. Heart J.* 34 (2013) 2949–3003, <https://doi.org/10.1093/eurheartj/ehz296>.
- [113] A. Kaplan, G. Amin, E. Abidi, R. Altara, G.W. Booz, F.A. Zoueini, Role of ranolazine in heart failure: from cellular to clinic perspective, *Eur. J. Pharmacol.* 919 (2022) 174787, <https://doi.org/10.1016/j.ejphar.2022.174787>.
- [114] L.S. Maier, B. Layug, E. Karwatowska-Prokopczuk, L. Belardinelli, S. Lee, J. Sander, C. Lang, R. Wachter, F. Edelmann, G. Hasenfuss, C. Jacobshagen, RANO-LazIne for the treatment of diastolic heart failure in patients with preserved ejection fraction: the RALI-DHF proof-of-concept study, *JACC Heart Fail.* 1 (2013) 115–122, <https://doi.org/10.1016/j.jchf.2012.12.002>.
- [115] G.L. Murray, J. Colombo, Ranolazine preserves and improves left ventricular ejection fraction and autonomic measures when added to guideline-driven therapy in chronic heart failure, *Heart Int.* 9 (2014) 66–73, <https://doi.org/10.5301/heartint.5000219>.
- [116] Y. Rabanal-Ruiz, E. Llanos-González, F.J. Alcaín, The use of coenzyme Q10 in cardiovascular diseases, *Antioxidants (Basel)* 10 (2021) 755, <https://doi.org/10.3390/antiox10050755>.
- [117] M. Jafari, S.M. Mousavi, A. Asgharzadeh, N. Yazdani, Coenzyme Q10 in the treatment of heart failure: a systematic review of systematic reviews, *Indian Heart J.* 70 (2018) S111–S117, <https://doi.org/10.1016/j.ihj.2018.01.031>.
- [118] S.A. Mortensen, F. Rosenfeldt, A. Kumar, P. Dolliner, K.J. Filipiak, D. Pella, U. Alehagen, G. Steurer, G.P. Littarru, Q-SYMBIO Study Investigators, The effect of coenzyme Q10 on morbidity and mortality in chronic heart failure: results from Q-SYMBIO: a randomized double-blind trial, *JACC Heart Fail.* 2 (2014) 641–649, <https://doi.org/10.1016/j.jchf.2014.06.008>.
- [119] J. Schwemmlin, C. Maack, E. Bertero, Mitochondria as therapeutic targets in heart failure, *Curr. Heart Fail. Rep.* 19 (2022) 27–37, <https://doi.org/10.1007/s11897-022-00539-0>.
- [120] M.A. Daubert, E. Yow, G. Dunn, S. Marchev, H. Barnhart, P.S. Douglas, C. O'Connor, S. Goldstein, J.E. Udelson, H.N. Sabbah, Novel mitochondria-targeting peptide in heart failure treatment: a randomized, placebo-controlled trial of elamipretide, *Circ. Heart Fail.* 10 (2017), <https://doi.org/10.1161/circheartfailure.117.004389>.
- [121] W. Yu, C. Chen, Y. Fu, X. Wang, W. Wang, Insulin signaling: a possible pathogenesis of cardiac hypertrophy, *Cardiovasc. Ther.* 28 (2010) 101–105, <https://doi.org/10.1111/j.1755-5922.2009.00120.x>.
- [122] J.J. Rayner, R. Banerjee, C.J. Holloway, A.J.M. Lewis, M.A. Peterzan, J. M. Francis, S. Neubauer, O.J. Rider, The relative contribution of metabolic and structural abnormalities to diastolic dysfunction in obesity, *Int. J. Obes.* 42 (2018) 441–447, <https://doi.org/10.1038/s41301-017-239>.
- [123] P. Geraldes, G.L. King, Activation of protein kinase C isoforms and its impact on diabetic complications, *Circ. Res.* 106 (2010) 1319–1331, <https://doi.org/10.1161/CIRCRESAHA.110.217117>.
- [124] I. Ozakca, A.T. Ozelikay, Roles of PKC isoforms in development of diabetes-induced cardiovascular complications, in: *Diabetic Cardiomyopathy*, Springer New York, New York, NY, 2014, pp. 269–284.
- [125] G. Jia, A.R. Aroor, J.R. Sowers, The role of mineralocorticoid receptor signaling in the cross-talk between adipose tissue and the vascular wall, *Cardiovasc. Res.* 113 (2017) 1055–1063, <https://doi.org/10.1093/cvr/cvx097>.
- [126] J. Butler, J.A. Ezekowitz, S.P. Collins, M.M. Givertz, J.R. Teerlink, M.N. Walsh, N. M. Albert, C.A. Westlake Canary, P.E. Carson, M. Colvin-Adams, J.C. Fang, A. F. Hernandez, R.E. Hersherberger, S.D. Katz, J.G. Rogers, J.A. Spertus, W. G. Stevenson, N.K. Sweitzer, W.H.W. Tang, W.G. Stough, R.C. Starling, Update on aldosterone antagonists use in heart failure with reduced left ventricular ejection fraction. Heart Failure Society of America Guidelines Committee, *J. Card. Fail.* 18 (2012) 265–281, <https://doi.org/10.1016/j.cardfail.2012.02.005>.
- [127] N.M. Albert, C.W. Yancy, L. Liang, X. Zhao, A.F. Hernandez, E.D. Peterson, C. P. Cannon, G.C. Fonarow, Use of aldosterone antagonists in heart failure, *JAMA.* 302 (2009) 1658–1665, <https://doi.org/10.1001/jama.2009.1493>.
- [128] S.-B. Catrina, X. Zheng, Hypoxia and hypoxia-inducible factors in diabetes and its complications, *Diabetologia.* 64 (2021) 709–716, <https://doi.org/10.1007/s00125-021-05380-z>.
- [129] D. Güzel, A.D. Dursun, H. Fiçıcılar, D. Tekin, A. Tanyeli, F. Akat, F. Topal Çelikkian, B. Sabuncuoğlu, M. Baştug, Effect of intermittent hypoxia on the cardiac HIF-1/VEGF pathway in experimental type 1 diabetes mellitus, *Anatol. J. Cardiol.* 16 (2016) 76–83, <https://doi.org/10.5152/akd.2015.5925>.
- [130] K. Wang, X. Dai, J. He, X. Yan, C. Yang, X. Fan, S. Sun, J. Chen, J. Xu, Z. Deng, J. Fan, X. Yuan, H. Liu, E.C. Carlson, F. Shen, K.A. Wintergerst, D.J. Conklin, P. N. Epstein, C. Lu, Y. Tan, Endothelial overexpression of metallothionein prevents diabetes-induced impairment in ischemia angiogenesis through preservation of HIF-1 α /SDF-1/VEGF signaling in endothelial progenitor cells, *Diabetes.* 69 (2020) 1779–1792, <https://doi.org/10.2337/db19-0829>.
- [131] M. Knapp, X. Tu, R. Wu, Vascular endothelial dysfunction, a major mediator in diabetic cardiomyopathy, *Acta Pharmacol. Sin.* 40 (2019) 1–8, <https://doi.org/10.1038/s41401-018-0042-6>.
- [132] E. Lespagnol, L. Dauchet, M. Pawlak-Chaouch, C. Balestra, S. Berthoin, M. Feelisch, M. Roustit, J. Boissière, P. Fontaine, E. Heyman, Early endothelial dysfunction in type 1 diabetes is accompanied by an impairment of vascular smooth muscle function: a meta-analysis, *Front. Endocrinol. (Lausanne)* 11 (2020) 203, <https://doi.org/10.3389/fendo.2020.00203>.
- [133] Y. Shi, P.M. Vanhoutte, Macro- and microvascular endothelial dysfunction in diabetes, *J. Diabetes* 9 (2017) 434–449, <https://doi.org/10.1111/1753-0407.12521>.
- [134] S. Epelman, P.P. Liu, D.L. Mann, Role of innate and adaptive immune mechanisms in cardiac injury and repair, *Nat. Rev. Immunol.* 15 (2015) 117–129, <https://doi.org/10.1038/nri3800>.
- [135] S.D. Prabhu, N.G. Frangogiannis, The biological basis for cardiac repair after myocardial infarction: from inflammation to fibrosis: from inflammation to fibrosis, *Circ. Res.* 119 (2016) 91–112, <https://doi.org/10.1161/CIRCRESAHA.116.303577>.
- [136] Regulating Inflammatory Cytokines in the Diabetic Heart, *Oxidative Stress in Heart Diseases*, n.d.
- [137] B. Feng, S. Chen, B. George, Q. Feng, S. Chakrabarti, miR133a regulates cardiomyocyte hypertrophy in diabetes, *Diabetes Metab. Res. Rev.* 26 (2010) 40–49, <https://doi.org/10.1002/dmrr.1054>.
- [138] Q.A. Hathaway, M.V. Pinti, A.J. Durr, S. Waris, D.L. Shepherd, J.M. Hollander, Regulating microRNA expression: at the heart of diabetes mellitus and the mitochondrion, *Am. J. Physiol. Heart Circ. Physiol.* 314 (2018) H293–H310, <https://doi.org/10.1152/ajpheart.00520.2017>.
- [139] B. Dai, H. Li, J. Fan, Y. Zhao, Z. Yin, X. Nie, D.W. Wang, C. Chen, MiR-21 protected against diabetic cardiomyopathy induced diastolic dysfunction by targeting gelsolin, *Cardiovasc. Diabetol.* 17 (2018) 123, <https://doi.org/10.1186/s12933-018-0767-z>.

Update

Life Sciences

Volume 334, Issue , 1 December 2023, Page

DOI: <https://doi.org/10.1016/j.lfs.2023.122168>



ELSEVIER

Contents lists available at ScienceDirect

Life Sciences

journal homepage: www.elsevier.com/locate/lifescie



Corrigendum

Corrigendum to “Current understanding of structural and molecular changes in diabetic cardiomyopathy” [Life Sciences 332 (2023) 122087]

Md Sayeed Akhtar^{a,*}, Sirajudeen S. Alavudeen^a, Asif Raza^b, Mohammad Tarique Imam^c, Ziyad Saeed Almalki^c, Fauzia Tabassum^{d,e}, Mir Javid Iqbal^f

^a Department of Clinical Pharmacy, College of Pharmacy, King Khalid University, Al-Fara, Abha 62223, Saudi Arabia

^b Department of Pharmacology, Penn State Cancer Institute, CH72, Penn State College of Medicine, Penn State Milton S. Hershey Medical Center, 500 University Drive, Hershey, PA 17033, USA

^c Department of Clinical Pharmacy, College of Pharmacy, Prince Sattam Bin Abdulaziz University, Al-Kharj 16273, Saudi Arabia

^d Department of Pharmacology, College of Dentistry and Pharmacy, Buraydah Private College, Al Qassim 51418, Saudi Arabia

^e Department of Pharmacology, Vision College, Ishbiliya, Riyadh 13226-3830, Saudi Arabia

^f Department of Pharmaceutical Sciences, Northeastern University, Boston, MA 02115, USA

The authors regret that one of the author's name has been wrongly written as Ziad Saeed Almalki, his correct name is Ziyad Saeed Almalki.

The author's name listed in this corrigendum are correct and

complete.

The authors would like to apologise for any inconvenience caused.

DOI of original article: <https://doi.org/10.1016/j.lfs.2023.122087>.

* Corresponding author at: Department of Clinical Pharmacy, College of Pharmacy, King Khalid University, Al-Fara, Abha 62223, Saudi Arabia.

E-mail address: mdhsuain@kku.edu.sa (M.S. Akhtar).

<https://doi.org/10.1016/j.lfs.2023.122168>

Available online 8 November 2023

0024-3205/© 2023 Published by Elsevier Inc.